REVIEW ARTICLE

microRNAs regulation and its role as biomarkers in diseases*

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Abstract	MicroRNAs (miRNAs), approximately 21 to 23 nucleotides (nt) in length, belong to a set of small non-coding RNA molecules that were not thought to be functional until the recent decades. miRNAs play important roles in many diseases such as various kinds of cancers and immune disorders. Many studies have focused on the relationship between miRNAs and diseases. miRNAs are significant mediators in human growth and development and in the genesis and development of diseases. Almost 30% of the activity of protein-coding genes is forecasted to be regulated by miRNAs in mammals, and some miRNAs are regarded as potential
Received: 4 August 2015 Revised: 2 September 2015 Accepted: 25 January 2016	therapeutic targets for various diseases. In this review, we outline some functions of miRNAs, especially those related to diseases. Key words: MicroRNA; cancer; disease; drug

According to the central dogma of molecular biology, DNA is transcribed into messenger RNA (mRNA), and proteins are synthesized from mRNA via translation. However, in the recent decade, the complexity of the transcriptome has been appreciated further. An increasing number of unknown non-coding RNA species, including small RNAs, small nuclear RNA (snRNA), and small nucleolar RNA (snoRNA), have been discovered. Small RNAs include microRNAs (miRNAs), small interfering RNAs (siRNAs), and PIWI-interacting RNA (piR-NAs). These diverse RNA species further our understanding of the regulation of DNA, RNA, and protein. Most non-coding RNA species play crucial roles in regulation (Table 1). The diverse regulatory functions of miRNAs in the cell cycle and in cell proliferation and differentiation have been identified by a large number of studies. This review outlines miRNA biogenesis and the essential roles of miRNA in various diseases, especially cancer. In addition, the diagnostic and therapeutic application of miR-NAs, e.g., as disease biomarkers, and the development of new drugs targeting miRNA are discussed.

microRNAs (miRNAs)

miRNAs (also known as small molecular RNAs), ap-

proximately 21 to 23 nucleotides in length, are RNA molecules that are widely distributed in eukaryotes; these miRNA can regulate gene expression. The evolution of miRNA is relatively conservative and it belongs to a class of non-coding RNAs that are transcribed from DNA but are not further translated into proteins. miRNAs target messenger RNA (mRNA) with a specific combination to inhibit gene transcription and play a significant role in the regulation of gene expression, cell cycle, biological development, etc. [1-2]. The biogenesis of miRNA has been extensively studied. miRNAs are initially transcribed from intragenic or intergenic regions by RNA polymerase II as variable-length transcripts, usually between 1 kb and 3 kb in size, called long primary RNAs (pri-microRNAs) [3-5]. The pri-microRNA is then processed by nuclear RNase III enzyme Drosha together with DGCR8 (DiGeorge syndrome critical region gene 8), which is a double-stranded RNA binding domain (dsRBD) partner of Drosha, into short -70-nucleotide RNA hairpin structures called precursor microRNAs (pre-microRNAs). This pre-microRNA hairpin is then exported out of the nucleus into the cytoplasm with the help of RanGTP-dependent Exportin 5^[6–7].

In the cytoplasm, the pre-microRNA is processed by Dicer, another RNase III enzyme, into a mature doublestranded miRNA of variable length (-22 nucleotides)

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40

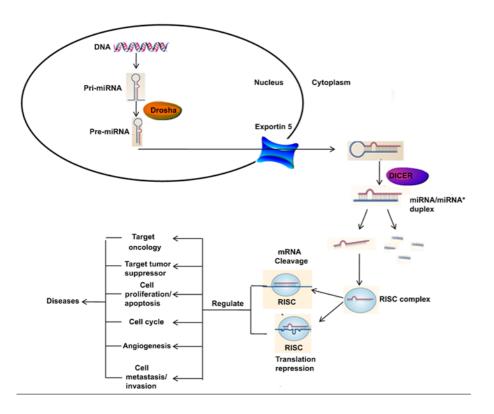
Table 1 The role of some non-coding RNA

tRNA (transfer RNA)	Translation	
siRNA (small interfering RNA)	RNA silencing	
microRNA (microRNA)	RNA silencing	
snRNA (small nuclear ribonucleic acid)	RNA splicing	
snoRNA (small nucleolar RNA)	Guide chemical modifications of other RNAs	
piRNA (Piwi-interacting RNA)	Gene silencing in retrotransposons and other genetic elements in germ line cells	
gRNA (guide RNA)	RNA editing	

^[8-9] (Fig. 1). TRBP, the dsRBD partner of Dicer, releases a 19–24-nucleotide fragment from the pre-microRNA hairpin. The miRNA plus-strand is usually degraded, and the separated strand of the mature miRNA is loaded onto the RNA-induced silencing complex (RISC), which is an effector complex comprising miRNA, Argonaute protein 1 to protein Argonaute 4, and other protein factors ^[10]. Argonaute protein contains a PIWI domain that binds to the 5' end of the miRNA; hence, it is important for the recognition of specific target mRNAs that result in post-transcriptional repression or degradation of the target mRNA depending on the pairing complementarities ^[11–12].

Discovery of miRNA

Victor Ambros and Gary Ruvkun discovered the first miRNA in 1993, when they were studying a gene named lin-4 in Caenorhabditis elegans; they confirmed that it controlled developmental timing in the worm by binding partially to the 3' untranslated region (UTR) of lin-14 mRNA ^[13–14]. In the following years, *lin-4* did not receive much attention and it was regarded an oddity in worm genetics, until a second small regulatory RNA, named let-7, was discovered in the worm ^[15]. These odd discoveries attracted the interests of Victor Ambros, David Bartel, and Thomas Tuschl; they began to look for other small RNAs in different organisms, and found a large number of miRNAs in C. elegans, Drosophila embryos, and human HeLa cells [16-17]. Subsequently, the first studies to report that miRNA dysregulation could cause human disease were published. Later, researchers found that miR-15a and miR-16-1 were deficient or downregulated in several cases of B cell chronic lymphocytic leukemia (CLL)^[18], which led to an increase in the number of studies evaluating the function of miRNAs. For example, miR-155, the first human oncogenic miRNA to be identified, was found to be upregulated in hematological malignancies and in inflammatory responses of macrophages [19-21]. miR-146a/b, miR-132, and miR-155 are upregulated in the human monocytic cell line THP-1 following stimulation by LPS ^[22]. Eventually, several studies investigating



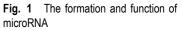


Table 2 Tissue-specfic miRNA expression signatures

Expression pattern	MicroRNA	
Enriched in brain	miR-12a, miR-125b, miR-128, miR-132,	
	miR-139, miR-7, miR-9, miR-153,	
	miR-124a, miR-124b, miR-135, miR-149,	
	miR-183, miR-190, miR-219	
Enriched in lung	miR-18, miR-19a, miR-24,	
	miR-32, miR-130, miR-213,	
	miR-20, miR-141, miR-193, miR-200b	
Enriched in spleen	miR-99a, miR-127, miR-142a, miR-142s,	
	miR-151, miR-189, miR-212	
Enriched in liver	miR-122a, miR-152, miR-194,	
	miR-199, miR-215	
Enriched in heart	miR-1b, miR-1d, miR-133, miR-206,	
	miR-208, miR-143	
Enriched in kidney	miR-30b, miR-30c, miR-18, miR-20,	
,	miR-24, miR-32, miR-141,	
	miR-193, miR-200b	
Enriched in	miR-181, miR-223, miR-142	
haematopoetic tissues		
Ubiquitously expressed	miR-16, miR-26a, miR-27a, miR-143a,	
	miR-21, let-7a, miR-7b,	
	miR-30b, miR-30c	

the different aspects and roles of miRNA were undertaken. Owing to these studies, we now have a deeper cognition of the functions of miRNA.

Distribution of miRNA

In the last 10 years, an increasing number of studies have focused on miRNA research and discovery. miRNAs are highly conserved and are ubiquitous in both animals and plants. Almost 30% of the activity of protein-coding genes is predicted to be regulated by miRNAs in mammals, and this aspect is being intensively researched upon in many fields. Different tissues have been found to exhibit distinctive patterns of miRNA expression. In addition, some miRNAs are ubiquitously expressed (Table 2)^[23].

miRNA and diseases

As seen in Fig. 1, miRNAs play pivotal roles in many biological processes such as cell growth, apoptosis, gene regulation, angiogenesis, cell cycle, and cancer cell metastasis/invasion. Therefore, miRNAs are associated with numerous human diseases. Dysregulation of miRNAs causes many diseases including various types of cancer and immune disorders. miRNAs can serve as promising therapeutic targets for several diseases because of their oncogenic functions, and potential therapies targeting miRNAs include miRNA silencing, antisense blocking, epigenetic modification, DNA copy number change, and genetic mutations^[24]. A useful strategy for tumor suppression involves the overexpression of miRNAs that suppress tumor growth and development. Thus, miRNAs are significant mediators of human physiology and disease and are crucial to early detection and prognosis of the disease and treatment-related decision-making.

miRNAs and cancer

Studies on miRNAs majorly focus on their potential role in tumor development. The function of miRNAs is similar to that of oncogenes or tumor suppressor genes, which are closely related to tumor development. First, research on worms and fruit flies confirmed that miR-NA function to regulate cell proliferation and apoptosis, which suggests that they are closely associated with hyperplastic diseases such as cancer. Second, many miRNA genes have been confirmed to be located in areas along with the variation of the tumor in the genome. Third, compared with normal tissue, tumor tissue or tumor cell lines exhibit widespread abnormal expression of regulatory miRNAs. Studies have found that mutations, deletions, and imbalance of post-transcriptional regulation, modification of DNA methylation in the promoter region of genes encoding miRNAs, and abnormal protein binding can cause abnormal expression of miRNAs.

Approximately more than 50% of human miRNA genes are located in cancer-associated regions or at chromosome fragile sites, which are susceptible to gene deletion, amplification, and mutations. In addition, abnormal expression of miRNA has been observed in many human cancers. Negative regulation of tumor growth by miRNAs has been observed in pancreatic cancer ^[25], breast cancer ^[26], prostate cancer ^[27], liver cancer ^[28], colon cancer ^[29–30], and ovarian cancer^[31]. Amplification of the miR-17-92 cluster in human B-cell lymphomas and upregulation of miR-155 in Burkitt's lymphoma have also been reported as examples of associations of oncogenic miRNA with human cancers ^[19, 32–33]. Different tumor tissues have markedly different miRNA expression spectra, with respect to the quantity and richness of miRNA expression. Another important aspect of the association of miRNAs with cancer is that the expression of some miRNAs is upregulated in some cancers, while in others, the expression of miR-NAs may be downregulated, which indicates that miR-NAs may be oncogenic or act as tumor suppressors. For example, in breast cancer, miR-10b $^{\scriptscriptstyle [34]}$, miR-21 $^{\scriptscriptstyle [35]}$, miR-22 [36], miR-27a [37], miR-155 [38], miR-210 [39], miR-221 [40], miR-222 [40], miR-328 [41], miR-373 [42], and miR-520c [42] were found to be upregulated, while let-7 [43], miR-7 [44], miR-9-1^[45], miR-17/miR-20^[46], miR-31^[47], miR-125a^[48], miR-125b [49], miR-146 [50], miR-200 family [51-52], miR-205 [52], miR-206 [53], and miR-335 [54] were found to be

downregulated. In chronic lymphocytic leukemia, miR-21 $^{\rm [55]}$ and miR-155 $^{\rm [55]}$ are upregulated, while miR-15 $^{\rm [18]},$ miR-16 [18], miR-29b [56], miR-29c [56], miR-34a [56], miR-143 [57], miR-145 [57], miR-181b [56], and miR-223 [56] are downregulated. In lung cancer, the miR-17-92 cluster, miR-21, miR-106a, and miR-155 are upregulated, while miR-1, the let-7 family, miR-7, miR-15a/miR-16, and the miR-29 family are downregulated [44, 58-61]. miR-221 and miR-222 are upregulated in prostate cancer, while the miR-15a-miR-16-1 cluster, miR-101, miR-127, and miR-449a are downregulated [32-44]. As for hepatocellular carcinoma, the miR-17-92 cluster, miR-21, miR-143, and miR-224 are upregulated, while miR-1, miR-101, and miR-122a are downregulated ^[28, 62-66]. Early studies in the field of neurobiology have shown that the tissue-specific abundant expression of miR-124 and miR-9 in the brain is regulated during brain development or during development of neurons and astrocytes in culture [67-68]. The spatial expression patterns of several miRNAs in human brain samples has revealed that miR-124 appears to be widely expressed in differentiated neurons, while miR-9 is more prominently expressed at an earlier stage in proliferating neuronal precursors [67, 69]. In addition, miRNA regulation has showed its significance in glioma development. miR-21 was found to be overexpressed in high-grade gliomas [70-71]. In addition, Chen [72] found that miR-107 inhibits glioma cell proliferation, migration, and invasion, and indicated that it could be a potential therapeutic target for glioma. Another study [73] showed that osthole could restrain the proliferation of human glioma cells and promote their apoptosis by upregulating the expression of miR-16 and downregulating the expression of MMP-9. Furthermore, through a study evaluating the expression of selected miRNAs (miR-16, -17, -19a, R-20a, -140, and -184) in an independent set of low-grade and secondary glioblastoma multiforme samples, the grade-associated regulation of these miRNAs was confirmed [74].

Role of miRNAs in the immune system

Since miRNAs are associated with the regulation of multiple genes, it is true for miRNAs involved in immune function. miRNAs have been shown to be associated with the proliferation of quiescent naïve T cells and effector T cells capable of differentiation that produce various cytokines during an effective immune response, as a result of their progressive differentiation owing to a marked change in gene expression profiles during and after infection, where they are expected to influence fundamental cellular processes ^[75–77]. Several miRNAs appear to be vital players in immunity. For example, miR-155, which has a specific role in inflammatory stress, has been identified as a key player in the biology of lymphocytes ^[21, 78]. miR-155 was found to be oncogenic after mice expressing

miR-155 in B cells developed lymphoma [79]. Moreover, miR-155 also plays a significant role in Alzheimer's disease by regulating T-cell functions during inflammation ^[80]. Another finding about miR-155 was that it could promote dendritic cell migration toward sites of ATP release, accompanied by inflammasome activation [81]. Members of the human miR-146 family have been identified as vital inflammatory inducers that regulate Toll-like receptor (TLR) signaling by a negative feedback mechanism ^[82]. miR-146a has been observed to regulate the innate immune response as a negative regulator of the expression of the NF-kB components IRAK1 and TRAF6, which encode key adapter molecules downstream of Toll-like receptors, by interfering with the NF-kB pathway [21-22, 83-84]. In addition, the miR-146 family was found to be involved in regulating lipid metabolism during inflammation through test the expression of the downstream factors of MyD88-Traf6 pathway, pro-inflammatory genes, after knocking down miR-146a and miR-146b expression ^[82]. miR-150 is highly upregulated during the development of mature T and B cells and is crucial to their terminal stages of differentiation [85-86]. miR-181 was preferentially expressed in B cells and its ectopic expression in hematopoietic progenitor cells during lineage differentiation led to a doubling of the number of cells of the B-lymphoid lineage [87].

miRNA-based clinical applications

Clinical research studies are currently focused on an increasing number of miRNAs. The miRNA-based classification of tumors seems to be more accurate than the mRNA expression profile-based classification of tumors ^[88]. Moreover, miRNAs could become useful tools in cancer diagnosis and prognosis and be effective therapeutic targets. Their applications in clinical practice mainly focus on two aspects: (1) First, their use as biomarkers of disease. miRNA profiles are potentially useful as early detection, classification, prognostic, and predictive biomarkers. As early detection biomarkers, they indicate the onset of a disease and often play a role in the disease. (2) Second, their use as attractive therapeutic targets. Various studies are underway worldwide to tap the potential of miRNAs for use as disease biomarkers. From http://www. clinicaltrials.gov/, a service of the U.S. National Institutes of Health, we found some pre-clinical trials evaluating the use of certain miRNAs as biomarkers of a particular disease (Table 3).

miRNAs have garnered considerable interest owing to their close association with many important diseases. As crucial targets for drug development, drugs corresponding to the miRNAs can be designed such that they achieve their therapeutic effect via the upregulation or downregulation of miRNAs or via silencing of miRNA expression. At present, molecular drug design based on miRNA is

Name of microRNA	Disease	Sponsor
MicroRNA 107	Alzheimer's disease	Shanghai Mental Health Center
Mir155	Biomarkers of sepsis (diagnostic and predictive value of circulating microRNAs during sepsis)	Changhai Hospital
Mir326	ESCC and NSCLC	China Medical University Hospital
Mir-29b	Oral squamous cell carcinoma	National Taiwan University Hospital
Mir-122	Chronic hepatitis C	National Taiwan University Hospital
Mir-29 family	Head-and-neck squamous cell carcinoma	National Taiwan University Hospital
Mir-10b	Astrocytoma; oligodendroglioma; oligoastrocytoma; anaplastic astrocytoma; anaplastic oligodendroglioma; anaplastic oligoastrocytoma; glioblastoma; brain tumors; brain cancer	National Taiwan University Hospital

Table 3 Pre-clinical trials of miRNAs as biomarkers used in diseases

still in its infancy. Many studies have mainly focused on simulating miRNAs to enhance the effectiveness of their role in targeting genes, or on designing small molecules as miRNA antagonists, such as miRNA antisense oligomeric nucleotides (anti-miRNA oligonucleotides, AMOs) and miRNA antagonism molecules (such as antagomirs).

The design of nucleic acid drugs in 2008 opened a new page of history in miRNA-based drug design. It should be noted that the first drug targeting miRNA was named miravirsen. The Danish pharmaceutical company Santaris Pharma announced that it would be the first to implement miRNA targets for drug clinical trials worldwide. Miravirsen is an antisense oligonucleotide with a locked nucleic acid (LNA)-modified oligonucleotide (SPC3649) complementary to miR-122. miR-122, expressed in the liver, is related to the replication of the hepatitis C virus (HCV) and to the regulation of cholesterol and lipid metabolism [89-90]. LNA nucleosides are a class of nucleic acid analogues in which an extra methylene bridge fixes the ribose moiety either in the C3'-endo or C2'-endo conformation. By locking the molecule, LNA oligonucleotides display unprecedented hybridization affinity towards complementary single-stranded RNA or double-stranded DNA ^[91–92]. In addition, they display excellent mismatch discrimination and high aqueous solubility. So-called LNA anti-miR constructs have been used successfully in several *in vitro* studies to knock down the expression of specific miRNAs ^[93-94]. According to the results of the phase I clinical trials of the drug, a dose-dependent effect of the drug on the reduction of HCV RNA levels was observed for an extended period, which was consistent with the results of the pre-clinical studies. According to the preliminary data obtained in phase II clinical trials, 18 individuals received miravirsen therapy, and six other individuals received a placebo treatment. No serious adverse reactions were observed, and the side effects included headache, diarrhea, and rhinitis; however, these side effects were moderate and infrequent [89].

Conclusion and future prospects

miRNAs have attracted considerable attention owing to their important role in cell differentiation, biological development, and in the development of various diseases. With further studies on the mechanisms underlying miR-NA function and on the relationships between miRNAs and diseases by using the latest high-throughput technology such as miRNAs chips, our understanding of the network of gene expression and regulation in eukaryotic cells will reach a new and higher level.

Various studies have found that miRNAs are significant mediators in human physiology and disease and are crucial to early detection and prognosis of the disease and treatment-related decision-making. Although the study of miRNAs has made great progress, especially with respect to the function of miRNAs in cell differentiation, gene regulation, and disease control, currently, the study of miRNAs is still in its nascent stage, with many genes yet to be identified and the mechanisms underlying the functions of most genes yet to be elucidated. As for the clinical applications, several technical hurdles to miRNA research remain, e.g., the cost of miRNA profiling and the development of drugs targeting miRNA is still high. Moreover, it is still technically difficult to achieve longterm and stable silencing of miRNA expression.

Conflicts of interest

The authors indicated no potential conflicts of interest.

References

- Hu XC, Yang JQ, Yang RJ, et al. MicroRNAs in tumor stem cells. Oncol Transl Med, 2015, 1: 92–96.
- Rodriguez A, Griffiths-Jones S, Ashurst JL, *et al.* Identification of mammalian microRNA host genes and transcription units. Genome Res, 2004, 14: 1902–1910.
- Landthaler M, Yalcin A, Tuschl T. The human DiGeorge syndrome critical region gene 8 and Its D. melanogaster homolog are required for miRNA biogenesis. Curr Biol, 2004, 14: 2162–2167.

- Lee Y, Jeon K, Lee JT, *et al.* MicroRNA maturation: stepwise processing and subcellular localization. EMBO J, 2002, 21: 4663–4670.
- Lee Y, Ahn C, Han J, et al. The nuclear RNase III Drosha initiates microRNA processing. Nature, 2003, 425: 415–419.
- Zeng Y, Cullen BR. Structural requirements for pre-microRNA binding and nuclear export by Exportin 5. Nucleic Acids Res, 2004, 32: 4776–4785.
- Bohnsack MT, Czaplinski K, Gorlich D. Exportin 5 is a RanGTP-dependent dsRNA-binding protein that mediates nuclear export of premiRNAs. RNA, 2004, 10: 185–191.
- Hutvagner G, Mclachlan J, Pasquinelli AE, et al. A cellular function for the RNA-interference enzyme Dicer in the maturation of the let-7 small temporal RNA. Science, 2001, 293: 834–838.
- Ketting RF, Fischer SE, Bernstein E, *et al.* Dicer functions in RNA interference and in synthesis of small RNA involved in developmental timing in C. elegans. Genes Dev, 2001, 15: 2654–2659.
- Chendrimada TP, Gregory RI, Kumaraswamy E, et al. TRBP recruits the Dicer complex to Ago2 for microRNA processing and gene silencing. Nature, 2005, 436: 740–744.
- Ma JB, Yuan YR, Meister G, et al. Structural basis for 5'-end-specific recognition of guide RNA by the A. fulgidus Piwi protein. Nature, 2005, 434: 666–670.
- Ghosh M, Meiss G, Pingoud A, *et al.* Structural insights into the mechanism of nuclease A, a betabeta alpha metal nuclease from Anabaena. J Biol Chem, 2005, 280: 27990–27997.
- Lee RC, Feinbaum RL, Ambros V. The C. elegans heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. Cell, 1993, 75: 843–854.
- Wightman B, Ha I, Ruvkun G. Posttranscriptional regulation of the heterochronic gene lin-14 by lin-4 mediates temporal pattern formation in C. elegans. Cell, 1993, 75: 855–862.
- Pasquinelli AE, Reinhart BJ, Slack F, *et al.* Conservation of the sequence and temporal expression of let-7 heterochronic regulatory RNA. Nature, 2000, 408: 86–89.
- Lagos-Quintana M, Rauhut R, Lendeckel W, et al. Identification of novel genes coding for small expressed RNAs. Science, 2001, 294: 853–858.
- Lau NC, Lim LP, Weinstein EG, *et al.* An abundant class of tiny RNAs with probable regulatory roles in Caenorhabditis elegans. Science, 2001, 294: 858–862.
- Calin GA, Dumitru CD, Shimizu M, *et al.* Frequent deletions and down-regulation of micro- RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. Proc Natl Acad Sci U S A, 2002, 99: 15524–15529.
- Eis PS, Tam W, Sun L, *et al.* Accumulation of miR-155 and BIC RNA in human B cell lymphomas. Proc Natl Acad Sci U S A, 2005, 102: 3627–3632.
- Costinean S, Zanesi N, Pekarsky Y, et al. Pre-B cell proliferation and lymphoblastic leukemia/high-grade lymphoma in E(mu)-miR155 transgenic mice. Proc Natl Acad Sci U S A, 2006, 103: 7024–7029.
- O'Connell RM, Taganov KD, Boldin MP, *et al.* MicroRNA-155 is induced during the macrophage inflammatory response. Proc Natl Acad Sci U S A, 2007, 104: 1604–1609.
- Taganov K D, Boldin MP, Chang KJ, et al. NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. Proc Natl Acad Sci U S A, 2006, 103: 12481–12486.
- Kusenda B, Mraz M, Mayer J, *et al.* MicroRNA biogenesis, functionality and cancer relevance. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub, 2006, 150: 205–215.

- Li M, Marin-Muller C, Bharadwaj U, *et al.* MicroRNAs: control and loss of control in human physiology and disease. World J Surg, 2009, 33: 667–684.
- Lee EJ, Gusev Y, Jiang J, *et al.* Expression profiling identifies microRNA signature in pancreatic cancer. Int J Cancer, 2007, 120: 1046–1054.
- Wang S, Huang J, Lyu H, et al. Functional cooperation of miR-125a, miR-125b, and miR-205 in entinostat-induced downregulation of erbB2/erbB3 and apoptosis in breast cancer cells. Cell Death Dis, 2013, 4: e556.
- Shi X B, Xue L, Yang J, et al. An androgen-regulated miRNA suppresses Bak1 expression and induces androgen-independent growth of prostate cancer cells. Proc Natl Acad Sci U S A, 2007, 104: 19983– 19988.
- Gramantieri L, Ferracin M, Fornari F, *et al.* Cyclin G1 is a target of miR-122a, a microRNA frequently down-regulated in human hepatocellular carcinoma. Cancer Res, 2007, 67: 6092–6099.
- Qin J, Wang F, Jiang H, *et al.* MicroRNA-145 suppresses cell migration and invasion by targeting paxillin in human colorectal cancer cells. Int J Clin Exp Patho, 2015, 8: 1328–1340.
- Tang Q, Zou Z, Zou C, *et al.* MicroRNA-93 suppress colorectal cancer development via Wnt/beta-catenin pathway downregulating. Tumour Biol, 2015, 36: 1701–1710.
- Sugio A, Iwasaki M, Habata S, et al. BAG3 upregulates Mcl-1 through downregulation of miR-29b to induce anticancer drug resistance in ovarian cancer. Gynecol Oncol, 2014, 134: 615–623.
- He L, Thomson JM, Hemann MT, et al. A microRNA polycistron as a potential human oncogene. Nature, 2005, 435: 828–833.
- Kluiver J, Poppema S, de Jong D, *et al.* BIC and miR-155 are highly expressed in Hodgkin, primary mediastinal and diffuse large B cell lymphomas. J Pathol, 2005, 207: 243–249.
- Eissa S, Matboli M, Shehata HH, et al. MicroRNA-10b and minichromosome maintenance complex component 5 gene as prognostic biomarkers in breast cancer. Tumour Biol, 2015, 36: 4487–4494.
- Toraih EA, Mohammed EA, Farrag S, *et al.* Pilot study of serum microRNA-21 as a diagnostic and prognostic biomarker in egyptian breast cancer patients. Mol Diagn Ther, 2015, 19: 179–190.
- Chen B, Tang H, Liu X, *et al.* miR-22 as a prognostic factor targets glucose transporter protein type 1 in breast cancer. Cancer Lett, 2015, 356: 410–417.
- Tang W, Zhu J, Su S, *et al.* MiR-27 as a prognostic marker for breast cancer progression and patient survival. PLoS One, 2012, 7: e51702.
- Shaker O, Maher M, Nassar Y, et al. Role of microRNAs -29b-2, -155, -197 and -205 as diagnostic biomarkers in serum of breast cancer females. Gene, 2015, 560: 77–82.
- Ivan M, Huang X. miR-210: fine-tuning the hypoxic response. Adv Exp Med Biol, 2014, 772: 205–227.
- Felicetti F, Errico MC, Bottero L, *et al.* The promyelocytic leukemia zinc finger-microRNA-221/-222 pathway controls melanoma progression through multiple oncogenic mechanisms. Cancer Res, 2008, 68: 2745–2754.
- Pan YZ, Morris ME, Yu AM. MicroRNA-328 negatively regulates the expression of breast cancer resistance protein (BCRP/ABCG2) in human cancer cells. Mol Pharmacol, 2009, 75: 1374–1379.
- Keklikoglou I, Koerner C, Schmidt C, et al. MicroRNA-520/373 family functions as a tumor suppressor in estrogen receptor negative breast cancer by targeting NF-kappaB and TGF-beta signaling pathways. Oncogene, 2012, 31: 4150–4163.
- 43. Yu F, Yao H, Zhu P, et al. let-7 regulates self renewal and tumorigenic-

ity of breast cancer cells. Cell, 2007, 131: 1109-1123.

- Webster RJ, Giles KM, Price KJ, et al. Regulation of epidermal growth factor receptor signaling in human cancer cells by microRNA-7. J Biol Chem, 2009, 284: 5731–5741.
- Lehmann U, Hasemeier B, Christgen M, et al. Epigenetic inactivation of microRNA gene hsa-mir-9-1 in human breast cancer. J Pathol, 2008, 214: 17–24.
- Calvano FC, Calvano-Mendes DC, Carvalho KC, et al. Triple-negative and luminal A breast tumors: differential expression of miR-18a-5p, miR-17-5p, and miR-20a-5p. Tumour Biol, 2014, 35: 7733–7741.
- Valastyan S, Reinhardt F, Benaich N, et al. A pleiotropically acting microRNA, miR-31, inhibits breast cancer metastasis. Cell, 2009, 137: 1032–1046.
- Nandy SB, Arumugam A, Subramani R, *et al.* MicroRNA-125a influences breast cancer stem cells by targeting leukemia inhibitory factor receptor which regulates the hippo signaling pathway. Oncotarget, 2015, 6: 17366–17378.
- Zhou M, Liu Z, Zhao Y, *et al.* MicroRNA-125b confers the resistance of breast cancer cells to paclitaxel through suppression of pro-apoptotic Bcl-2 antagonist killer 1 (Bak1) expression. J Biol Chem, 2010, 285: 21496–21507.
- Hurst DR, Edmonds MD, Scott GK, et al. Breast cancer metastasis suppressor 1 up-regulates miR-146, which suppresses breast cancer metastasis. Cancer Res, 2009, 69: 1279–1283.
- Castilla MA, Diaz-Martin J, Sarrio D, et al. MicroRNA-200 family modulation in distinct breast cancer phenotypes. PLoS One, 2012, 7: e47709.
- Paterson EL, Kolesnikoff N, Gregory PA, et al. The microRNA-200 family regulates epithelial to mesenchymal transition. Scientific World J, 2008, 8: 901–904.
- Kondo N, Toyama T, Sugiura H, et al. miR-206 Expression is downregulated in estrogen receptor alpha-positive human breast cancer. Cancer Res, 2008, 68: 5004–5008.
- Tavazoie SF, Alarcon C, Oskarsson T, *et al*. Endogenous human microRNAs that suppress breast cancer metastasis. Nature, 2008, 451: 147–152.
- Fulci V, Chiaretti S, Goldoni M, et al. Quantitative technologies establish a novel microRNA profile of chronic lymphocytic leukemia. Blood, 2007, 109: 4944–4951.
- Visone R, Rassenti LZ, Veronese A, *et al.* Karyotype-specific microRNA signature in chronic lymphocytic leukemia. Blood, 2009, 114: 3872–3879.
- Akao Y, Nakagawa Y, Kitade Y, *et al.* Downregulation of microRNAs-143 and -145 in B-cell malignancies. Cancer Sci, 2007, 98: 1914– 1920.
- Takamizawa J, Konishi H, Yanagisawa K, et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. Cancer Res, 2004, 64: 3753–3756.
- Bandi N, Zbinden S, Gugger M, et al. miR-15a and miR-16 are implicated in cell cycle regulation in a Rb-dependent manner and are frequently deleted or down-regulated in non-small cell lung cancer. Cancer Res, 2009, 69: 5553–5559.
- Fabbri M, Garzon R, Cimmino A, *et al.* MicroRNA-29 family reverts aberrant methylation in lung cancer by targeting DNA methyltransferases 3A and 3B. Proc Natl Acad Sci U S A, 2007, 104: 15805– 15810.
- Nasser MW, Datta J, Nuovo G, *et al.* Down-regulation of micro-RNA-1 (miR-1) in lung cancer. Suppression of tumorigenic property of lung cancer cells and their sensitization to doxorubicin-induced apoptosis by miR-1. J Biol Chem, 2008, 283: 33394–33405.

- Wang Y, Lee A T, Ma J Z, *et al.* Profiling microRNA expression in hepatocellular carcinoma reveals microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific target. J Biol Chem, 2008, 283: 13205–13215.
- Connolly E, Melegari M, Landgraf P, et al. Elevated expression of the miR-17-92 polycistron and miR-21 in hepadnavirus-associated hepatocellular carcinoma contributes to the malignant phenotype. Am J Pathol, 2008, 173: 856–864.
- Zhang X, Liu S, Hu T, *et al.* Up-regulated microRNA-143 transcribed by nuclear factor kappa B enhances hepatocarcinoma metastasis by repressing fibronectin expression. Hepatology, 2009, 50: 490–499.
- Su H, Yang J R, Xu T, *et al.* MicroRNA-101, down-regulated in hepatocellular carcinoma, promotes apoptosis and suppresses tumorigenicity. Cancer Res, 2009, 69: 1135–1142.
- Datta J, Kutay H, Nasser MW, *et al.* Methylation mediated silencing of MicroRNA-1 gene and its role in hepatocellular carcinogenesis. Cancer Res, 2008, 68: 5049–5058.
- Smirnova L, Grafe A, Seiler A, *et al*. Regulation of miRNA expression during neural cell specification. Eur J Neurosci, 2005, 21: 1469–1477.
- Sempere LF, Freemantle S, Pitha-Rowe I, *et al.* Expression profiling of mammalian microRNAs uncovers a subset of brain-expressed microRNAs with possible roles in murine and human neuronal differentiation. Genome Biol, 2004, 5: R13.
- Nelson PT, Baldwin DA, Kloosterman WP, et al. RAKE and LNA-ISH reveal microRNA expression and localization in archival human brain. RNA, 2006, 12: 187–191.
- Tomaselli S, Galeano F, Alon S, *et al.* Modulation of microRNA editing, expression and processing by ADAR2 deaminase in glioblastoma. Genome Biol, 2015, 16: 5.
- Wang P, Liu YH, Yao YL, *et al.* Long non-coding RNA CASC2 suppresses malignancy in human gliomas by miR-21. Cell Signal, 2015, 27: 275–282.
- Chen L, Li ZY, Xu SY, *et al.* Upregulation of miR-107 inhibits glioma angiogenesis and VEGF expression. Cell Mol Neurobiol, 2016, 36: 113–120.
- Lin K, Gao Z, Shang B, et al. Osthole suppresses the proliferation and accelerates the apoptosis of human glioma cells via the upregulation of microRNA-16 and downregulation of MMP-9. Mol Med Rep, 2015, 12: 4592–4597.
- Luo JW, Wang X, Yang Y, et al. Role of micro-RNA (miRNA) in pathogenesis of glioblastoma. Eur Rev Med Pharmacol Sci, 2015, 19: 1630–1639.
- Wu H, Neilson J R, Kumar P, et al. miRNA profiling of naive, effector and memory CD8 T cells. PLoS One, 2007, 2: e1020.
- 76. Starr TK, Jameson SC, Hogquist KA. Positive and negative selection of T cells. Annu Rev Immunol, 2003, 21: 139–176.
- Neilson JR, Zheng GX, Burge CB, *et al.* Dynamic regulation of miRNA expression in ordered stages of cellular development. Genes Dev, 2007, 21: 578–589.
- Ji Y, Wrzesinski C, Yu Z, *et al.* miR-155 augments CD8+ T-cell antitumor activity in lymphoreplete hosts by enhancing responsiveness to homeostatic gammac cytokines. Proc Natl Acad Sci U S A, 2015, 112: 476–481.
- Mashima R. Physiological roles of miR-155. Immunology, 2015, 145: 323–333.
- Song J, Lee JE. miR-155 is involved in Alzheimer's disease by regulating T lymphocyte function. Front Aging Neurosci, 2015, 7: 61.
- Chen S, Smith BA, Iype J, et al. MicroRNA-155-deficient dendritic cells cause less severe GVHD through reduced migration and defec-

tive inflammasome activation. Blood, 2015, 126: 103-112.

- Ordas A, Kanwal Z, Lindenberg V, et al. MicroRNA-146 function in the innate immune transcriptome response of zebrafish embryos to Salmonella typhimurium infection. BMC Genomics, 2013, 14: 696.
- O'Neill LA. 'Fine tuning' TLR signaling. Nat Immunol, 2008, 9: 459– 461.
- Arron JR, Walsh MC, Choi Y. TRAF-mediated TNFR-family signaling. Curr Protoc Immunol, 2002, 11: 11–19.
- Xiao C, Calado DP, Galler G, *et al.* MiR-150 controls B cell differentiation by targeting the transcription factor c-Myb. Cell, 2007, 131: 146–159.
- Garcia P, Frampton J. Hematopoietic lineage commitment: miRNAs add specificity to a widely expressed transcription factor. Dev Cell, 2008, 14: 815–816.
- Chen CZ, Li L, Lodish HF, et al. MicroRNAs modulate hematopoietic lineage differentiation. Science, 2004, 303: 83–86.
- Lu J, Getz G, Miska EA, *et al.* MicroRNA expression profiles classify human cancers. Nature, 2005, 435: 834–838.
- 89. Janssen HL, Reesink HW, Lawitz EJ, et al. Treatment of HCV infec-

tion by targeting microRNA. N Engl J Med, 2013, 368: 1685-1694.

- Lindow M, Kauppinen S. Discovering the first microRNA-targeted drug. J Cell Biol, 2012, 199: 407–412.
- Sonkoly E, Wei T, Janson PC, et al. MicroRNAs: novel regulators involved in the pathogenesis of psoriasis?. PLoS One, 2007, 2: e610.
- Vester B, Wengel J. LNA (locked nucleic acid): high-affinity targeting of complementary RNA and DNA. Biochemistry, 2004, 43: 13233– 13241.
- Bottoni A, Piccin D, Tagliati F, et al. miR-15a and miR-16-1 down-regulation in pituitary adenomas. J Cell Physiol, 2005, 204: 280–285.
- 94. Johnson SM, Grosshans H, Shingara J, *et al.* RAS is regulated by the let-7 microRNA family. Cell, 2005, 120: 635–647.

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